

Searching for Silver Bullets

An Alternative Strategy for Crystallizing Macromolecules

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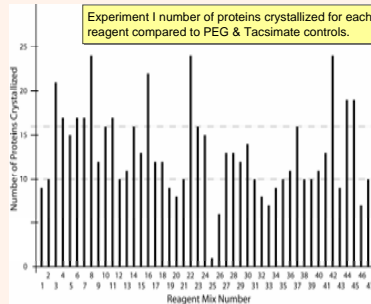
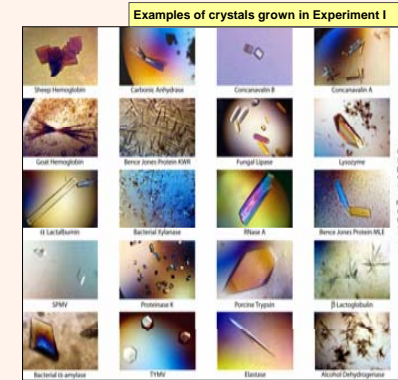
What we did

Based on a hypothesis that various small molecules might establish stabilizing, intermolecular, non covalent crosslinks in protein crystals and thereby promote lattice formation, we carried out three separate experiments. We assessed the impact of 200 chemicals on the propensity of 81 different proteins and viruses to crystallize. The experiments were comprised of 18,240 vapor diffusion trials. A salient feature of the experiments was that, aside from the inclusion of the reagent mixes, only two base crystallization conditions were used, 30% PEG 3350, and 50% Tacsimate™, both buffered at pH 7.

How we did it

- All reagents were:
 - Manually dispensed sitting drop vapor diffusion
 - Incubated at 25° Celsius
 - Reviewed manually at 20 and 200x using polarized light at 4, 8 and 12 weeks
 - Expt I used 3 different reservoir concentrations, Expt II used 2, and Expt III used 1
- All experiments used two fundamental conditions:
 - 30% w/v PEG 3,350, 0.1 M Hepes pH 7.0
 - 50% Tacsimate™ pH 7.0
- A total of 200 chemicals compounds were tested.
- Screened a total of 81 total proteins in all 3 expts: Expt I=60; Expt II=67; Expt III=66
- Expt I & II used 48 different reagent mixes, 2 of which were PEG and Tacsimate controls. The remaining 46 reagents were comprised of 3 to 20 different chemicals, on average 3 to 6 chemicals in each reagent.
- Expt III used 24 different bioactive compound reagent mixes, 1 of which was a PEG control. Each reagent is comprised of 4 to 5 bioactive compounds such as coenzymes, prosthetic groups, inhibitors, drugs, effectors, nucleotides, amino acids/peptides, and sugars/oligosaccharides. Sample arrays set in duplicate.
- A total of 18,240 experiment drops were set for three groups of experiments.

Experimental Results

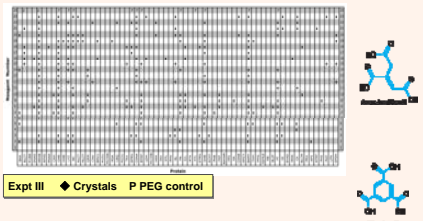
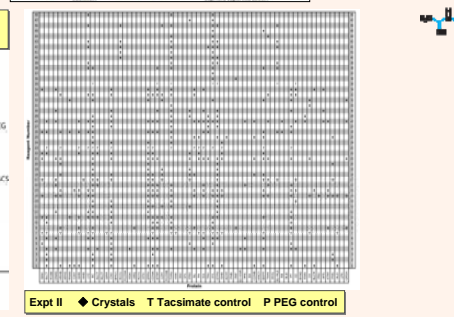
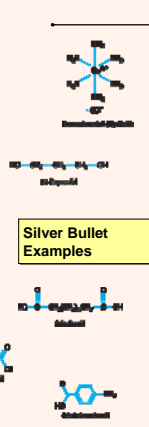
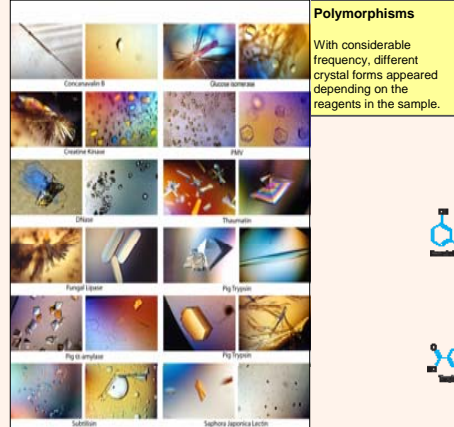


Experiment I

1. spermidine, spermine, cadaverine, putrescine, 1,8 diamino octane
2. 6-amino hexanoic acid, 3-aminopropionic acid, 4-aminobutyric acid
3. oxamic acid, sulfanilic acid, 4-aminobenzoic acid
4. Mix #2 and Mix #3
5. glutamic acid, hexadecanoic acid, dodecanoic acid, fumaric acid
6. oxalic acid, maleic acid, oxaloacetic acid, terephthalic acid
7. Mix #5 and Mix #6
8. Pentaglycine, triglycine
9. poly-L-lysine, poly-L-ornithine, poly-DL-alanine
10. poly-L-glutamic acid, poly-DL-alanine
11. Mix #8 and Mix #10
12. Protamine
13. heparin, dextran sulfate
14. dipA4, dipA6
15. dipT4, dipT2
16. pyrophosphate, tetraphosphate
17. ornithine, arginine, asparagine, glutamine
18. Tacsimate, PEG 3350
19. glycerol, sucrose, sorbitol
20. TMAO, pectine
21. sarcosine, glutamate, glycine, betaine
22. hexanoic acid, glyoxylic acid, isobutyric acid, D-hydroxyphenylacetic acid
23. palmitic acid, octanoic acid, stearoylamine
24. butyric acid, isosorbitol, glycerol-2-phosphate
25. potassium acetate, goldodium chloride, cobalt hexamine
26. hexadecyl trimethylammonium bromide
27. Tacsimate, Mix #8
28. Tacsimate, poly-L-ornithine, lysine, glutamic acid, alanine
29. Tacsimate, Mix #2
30. Tacsimate, Mix #3
31. Tacsimate, Mix #5
32. Tacsimate, Mix #5
33. Tacsimate, Mix #1
34. Tacsimate, Mix #19
35. Tacsimate, Mix #17
36. Tacsimate, Mix #13
37. PEG 3350
38. Tacsimate, dipA4, dipA6
39. Tacsimate
40. Tacsimate, dipT12, dipT4
41. Mix #8, Mix #9
42. Mix #8, Mix #9
43. Mix #1, Mix #3
44. Mix #1, Mix #3
45. The 20 amino acids
46. Tacsimate, the 20 amino acids
47. Tacsimate, the 20 amino acids
48. Tacsimate, proline

Experiment II

1. Schardinger's dextrin
2. NDSB-201
3. NDSB-195, NDSB-201, NDSB-211, NDSB-221, NDSB-256
4. tauric acid
5. streptomycin
6. trimelic acid, pyrophosphoric acid
7. tetraphosphoric acid, cobalt hexamine
8. maleic acid, myoinositol, phytic acid
9. Mix #8 and Mix #8
10. piperidine ethane sulfonic acid, anthraquinone disulfonic acid
11. neosporin, tetrahydroborate
12. pyromellitic acid, 2,2-thiodiglycolic acid, barbituric acid, terephthalic acid
13. fumaric acid, maleic acid, glutaric acid, pimelic acid, succinic acid
14. suberic acid, sebacic acid, hexadecanoic acid, dodecanoic acid
15. fumaric acid, maleic acid, sebacic acid, hexadecanoic acid
16. maleic acid, glutaric acid, succinic acid, sebacic acid, dodecanoic acid, oxamic acid
17. malonase, sucrose, melibiose, maltose, cellobiose
18. arabinose, maltotriose, melitriose
19. melibiose, stachyose
20. cyclohexan, 2-hydroxypropyl-β-cyclodextrin, 2,6-di-O-methyl-β-cyclodextrin
21. D, L, tetra, pentaglycine
22. 1,3-bisphosphoric acid, 1,2-hydroxyethyl, MPD, 1,6-hexanediol
23. PEG 3350
24. hexammine, 1,4-diaminobutane, spermine, 1,8-diaminooctane, cadaverine
25. pyrophosphate, phosphorous acid triethyl ester, phytic acid
26. Mix #4 and Mix #10
27. NDSB-201 and Mix #13
28. Mix #15 and Mix #16
29. phenyl urea, sodium L-pentanesulfonate, sulfanilic acid, salicin, pentamercaptoacetic acid
30. oxamic acid, fumarate, fumaramide, putrescine, pentane tetracarboxylic acid
31. succinic acid, PAA, sulfonic acid, 3-oxobutyric acid, pentane tetracarboxylic acid
32. 1-thioethylglycine, Anapoe-309, MEGA-9, zonyl-β-D-glucoside
33. streptic acid, maleic acid, pyromellitic acid, terephthalic acid
34. phloroglucinol, phytic acid, anthraquinone disulfonic acid, barbituric acid, tetraphosphoric acid
35. Tacsimate, Mix #17
36. Tacsimate and Mix #2
37. Tacsimate and Mix #5
38. Tacsimate and Mix #9
39. Tacsimate and Mix #10
40. Tacsimate and Mix #12
41. Tacsimate and Mix #13
42. Tacsimate and Mix #14
43. Tacsimate and Mix #15
44. Mix #1, Mix #3
45. Tacsimate and Mix #20
46. Tacsimate and Mix #20
47. Tacsimate and Mix #24
48. Tacsimate and Mix #21



Experiment III - Bioactive Compounds

1. S-adenosylmethionine, β-glycerophosphate, stachyose
2. levon, phosphoric acid, pyruvic acid, melitriose
3. FAD, phosphoglyceric acid, spermine
4. rhythm, pyrophosphate, glyoxylic acid, Schardinger's dextrin
5. NAD, hexose-1,6-diphosphate, spermidine
6. pyruvoyl-β-phosphate, pyruvic acid, cyclohexan
7. ADP, AMP, GMP
8. GMP, GDP, nucleoside acid
9. GTP, cTMP, chaperoninase
10. ATP, cAMP, estradiol, galactose
11. Tacsimate
12. spic acid, creatine, glutathione, parathionic acid
13. acetoamine, bay lactic acid, glucose
14. levon mononucleotide, trolean, acetyl choline
15. propanol A, GMP, fumaric acid, glucose
16. ADP, tetrahydrolic acid, succinic acid, glucose
17. adonin, phosphoryl choline, raffinose
18. GMP, cholesteryl, rhythm, oxamic acid
19. phosphocreatine, benzylarginine ethyl ester, phenobarbital, tetraethylene
20. ATP, hexameth, melitriose, linopirin
21. phosphorylthiose-1-phosphate, maleic acid, n-acetyl-D-glucosamine
22. UMP, ribose, phytic acid, palmitic acid
23. The 20 amino acids
24. PEG 3350

Positive charge centers, Diols, Dicarboxylic acids, Mixed amino and carboxylic acids, Polar amino acids, Glycine oligomers, Organic acids, Charge symmetric, and Diamines

Condensation of Statistics

	Experiment			
	All	I	II	III
How many proteins were investigated?	81	60	67	66
How many proteins crystallized?	65	48	50	50
How many proteins crystallized in Polyethylene glycol controls?	20	11	14	13
in Tacsimate controls?	6	5	4	N/A
in both controls?	5	5	3	N/A
Total controls	31	21	21	13
How many proteins crystallized only in the presence of some reagent mix?	35	28	29	37
How many reagents exceeded the Polyethylene glycol controls?	11	7	4	4
Tacsimate controls?	7	1	N/A	
How many proteins that did NOT crystallize in controls, crystallized in ONLY				
One reagent mix?	4	7	13	
Two reagent mixes?	2	4	9	
Three reagent mixes?	1	4	6	

What happened

Overall, 65 proteins (85%) were crystallized. Most significant was that 35 of the 65 (54%) crystallized only in the presence of one or more reagent mixes, but not in control samples lacking any additives. Among the most promising types of reagent mixes were those composed of polyvalent, charged groups, such as di and tri carboxylic acids, diamino compounds, molecules bearing one or more sulfonyl or phosphate groups, and a broad range of common biochemicals, coenzymes, biological effectors, and ligands. Less promising reagent mixes were composed of osmolites, polyamines, detergents and sugars.

Summary

We propose that an alternate approach to crystallizing proteins might be developed, which employs a limited set of fundamental crystallization conditions combined with a broad screen of potentially useful small molecule additives.

What's next

The evaluation of more reagent mixes, the influence of pH and the use of common dehydrants.

Thank you Aaron Greenwood, Joe Luft & Peter Nguyen.